the endosafe® nexgen-pts™

GRAM ID ASSAY GUIDE

Gram ID Test on the nexgen-PTS™

A routine Gram ID assay is conducted by preparing an isolate and following the simple prompts on the nexgen-PTS™ instrument. The following represents a typical assay procedure:

- Isolate necessary colonies prior to testing, approximately 24 hours depending on the isolate.
- The nexgen-PTS™ Gram ID offers 2 assay report formats:
  - One assay result format will report the presence of either Gram-negative microorganisms, yeast/mold, or Gram-positive microorganisms in approximately 7 minutes.
  - The second assay result format will report the presence of Gram-negative and Gram-positive in approximately 3 minutes (this format does not differentiate between Gram-positive and yeast/mold isolates).
- Refer to the Certificate of Analysis for the calibration code to be used for each assay result format.

Sample Preparation

- From an isolated culture, carefully remove colonies from agar surface and suspend cells in approximately 2 mL of LPS-free saline or LRW and adjust to a 0.5 McFarland Equivalence Turbidity Standard.
- Up to 4 samples can be run at one time. A new pipette tip should be used for each sample added.

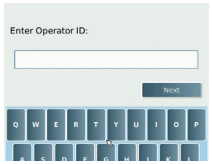
Note: Do not use cotton tip swabs of any kind during sample preparation.

Necessary Accessories for Running a Gram ID Test on the nexgen-PTS™*

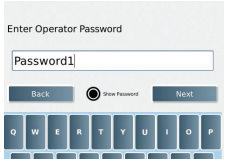
- Isolated culture of desired sample(s) to be tested.
- 0.5 MacFarland Equivalence Turbidity Standard. Compare the isolated colonies in solution to the MacFarland Standard.
- 0.9% saline or LAL reagent water
- Sterile inoculating loops
- Pipettor and sterile tips. The tips should not contain a cotton plug or filter.
- Disposable, endotoxin-free glass dilution tubes or sterile, disposable polystyrene tubes

** Not Included. See catalog for pricing and product codes.*

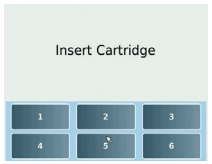
01 Turn on the reader by pressing the button at the rear right-hand side of the back panel. As the instrument boots up, the Charles River logo will display on the touch screen.



02 At the **Enter Operator ID** prompt, enter your username and click **Next**.



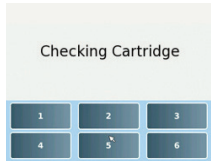
03 Login is only required if user management is switched on.



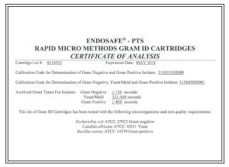
04 As the reader warms in preparation for use, the **Initializing** message appears. When the unit has reached 37.0 °C, the screen will display the **Insert Cartridge** message.

05 Allow cartridges to reach room temperature in pouch before using. Remove cartridge from pouch and avoid touching the sample wells.

06 Insert the cartridge firmly into the slot at the front of the reader with sample wells facing up.



07 The reader will then verify cartridge integrity; used or damaged cartridges will signal an alert.



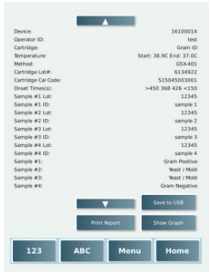
08 Prior to testing, be sure you have access to the **Certificate of Analysis** (COA), which can be found within your Endosafe®-PTS™ cartridge package or on the Endosafe® customer portal (www.criver.com/endosafeportal). This information includes the cartridge lot number and calibration code.

09 A series of prompts will appear for you to enter the information necessary to run the assay, including **Sample ID** and **Sample Lot** for each sample (up to 4 at a time). Enter values for each field, clicking the **Next** button until all fields are complete.

10 Review information added to ensure it is correct before clicking the **Next** button.

11 Add sample to cartridge. When adding each sample to the cartridge, use a new tip for each sample. Press **Done** – **Start Test** when ready. Be sure to dispense exactly 25 μL of sample into each well and to hold the tip at an angle and not completely at the bottom of the sample reservoir.

12 Test results will appear on screen when the assay is complete. You can send this data to a network printer, Zebra printer, download to Charles River Cortex™, or save to a USB.



13 Results will be reported based on the calibration code used and the reaction times of each sample added. One calibration code from the Certificate of Analysis will provide results in either Gram-negative or Gram-positive, where the other calibration code determines Gram-negative, Gram-positive, and yeast/mold. See the Certificate of Analysis for the calibration code necessary for the specific lot of cartridges being used.

charles river

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